FILE	'MEDLINE,	EMBASE, BIOSIS' ENTERED AT 12:58:08 ON 22 AUG 2005
L1	12777	S MENDEZ?/AU OR FINER?/AU
L2	10971	S (SHUTTLE OR HYBRID) (S) (VECTOR OR PLASMID OR CONSTRUCT)
L3	463478	S MARKER OR SELECTABLE
L4	15458	S ADENOVRIUS OR AAV OR "ADENO ASSOCIATED" OR POX OR PAPOVA OR
L5	20758	S ATTT OR TN7 OR FLP OR LOX OR CRE OR CIRCULARIZATION
L6	7	S L1 AND L2
L7	3	DUP REM L6 (4 DUPLICATES REMOVED)
L8	2	S L7 NOT PY>=2000
L9	1	S L2 AND L3 AND L4
L10	1033	S L5 AND L3
L11	15	S L10 AND L2
L12	2	S L11 AND YEAST
L13	7	DUP REM L11 (8 DUPLICATES REMOVED)
L14	1	DUP REM L12 (1 DUPLICATE REMOVED)
L15	4	S L13 NOT PY>=2000

L9 ANSWER 1 OF 1 BIOSIS COPYRIGHT (c) 2005 The Thomson Corporation on STN

ACCESSION NUMBER: 2004:289891 BIOSIS DOCUMENT NUMBER: PREV200400288085

TITLE: Transfection-free and scalable recombinant AAV

vector production using HSV/AAV hybrids.

AUTHOR(S): Booth, M. J.; Mistry, A.; Li, X.; Thrasher, A.; Coffin, R.

S. [Reprint Author]

CORPORATE SOURCE: Windeyer InstDept Immunol and Mol Pathol, Univ Coll London,

46 Cleveland St, London, W1T 4JF, England

Gene Therapy, (May 2004) Vol. 11, No. 10, pp. 829-837.

print.

ISSN: 0969-7128 (ISSN print).

DOCUMENT TYPE: Article LANGUAGE: English

SOURCE:

ENTRY DATE: Entered STN: 16 Jun 2004

Last Updated on STN: 16 Jun 2004

AB Adeno-associated virus (AAV) vectors are

highly efficient tools for use in gene therapy. Current production methods rely on plasmid transfection and are not generally considered amenable to scale-up. To improve recombinant AAV (rAAV) vector production in terms of both final titre and simplicity, we constructed recombinant herpes simplex virus (HSV) vectors, either disabled (ICP27 deleted) or nondisabled, encoding the AAV rep and cap genes. We also integrated an rAAVGFP construct into the nondisabled vector and also into a second pair of HSV vectors (disabled and nondisabled) not expressing rep and cap. Transgene incorporation and expression was confirmed by Southern and Western blot, respectively. Optimal double-infection ratios were established for disabled and nondisabled pairs of rep/cap-expressing and rAAVGFP-containing vectors, resulting in up to 1.55 x 1012 rAAV capsids and 4 x 108 expression units from approximately 1 x 107 BHK producer cells. Functionality of the prepared vector was confirmed by the detection of abundant green fluorescent protein (GFP) expression following injections of rAAV preparations into the rat brain. This paper therefore describes a simple, efficient, and transfection-free rAAV production process based on the use of HSV and not relying on a proviral cell line that, with appropriate scale-up, could yield quantities of rAAV sufficient for routine clinical use.

MEDLINE on STN L14 ANSWER 1 OF 1 DUPLICATE 1

ACCESSION NUMBER: 88210531 MEDLINE DOCUMENT NUMBER: PubMed ID: 3449223

Terminal segment of Kluyveromyces lactis linear DNA TITLE:

plasmid pGKL2 supports autonomous replication of

hybrid plasmids in Saccharomyces cerevisiae.

Fujimura H; Hishinuma F; Gunge N AUTHOR:

Mitsubishi Kasei Institute of Life Sciences, Tokyo, Japan. CORPORATE SOURCE:

Current genetics, (1987) 12 (2) 99-104. SOURCE:

Journal code: 8004904. ISSN: 0172-8083.

PUB. COUNTRY: United States

Journal; Article; (JOURNAL ARTICLE) DOCUMENT TYPE:

LANGUAGE: English

FILE SEGMENT: Priority Journals .

OTHER SOURCE: GENBANK-X05601

198806 ENTRY MONTH:

=>

ENTRY DATE: Entered STN: 19900308

Last Updated on STN: 19990129 Entered Medline: 19880617

AΒ By use of linear DNA plasmid pGKL2 from the yeast Kluyveromyces lactis we have constructed hybrid plasmids carrying a LEU2 gene of Saccharomyces cerevisiae as a selectable marker. The replication properties of hybrid plasmids in yeasts were investigated. We demonstrated that the insertion of a LEU2 gene into pGKL2 resulted in circularization of the hybrid plasmids and pGKL2 segment supported autonomous replication of the plasmids. Moreover, the hybrid plasmids propagated autonomously, independently of the presence of the natural pGKL2 plasmid.

L15 ANSWER 1 OF 4 MEDLINE on STN ACCESSION NUMBER: 1999200479 MEDLINE

DOCUMENT NUMBER:

PubMed ID: 10102356

TITLE:

pBECKS2000: a novel plasmid series for the facile creation of complex binary vectors, which incorporates "clean-gene"

facilities.

AUTHOR:

McCormac A C; Elliott M C; Chen D F

CORPORATE SOURCE:

The Norman Borlaug Institute for Plant Science Research, De

Montfort University, Scraptoft, Leicester, UK..

amccorma@dmu.ac.uk

SOURCE:

Molecular & general genetics: MGG, (1999 Mar) 261 (2)

226-35.

Journal code: 0125036. ISSN: 0026-8925. GERMANY: Germany, Federal Republic of Journal; Article; (JOURNAL ARTICLE)

LANGUAGE:

English

FILE SEGMENT:

PUB. COUNTRY:

DOCUMENT TYPE:

Priority Journals

ENTRY MONTH:

199904

ENTRY DATE:

Entered STN: 19990511

Last Updated on STN: 19990511 Entered Medline: 19990426

A new plasmid series has been created for Agrobacterium-mediated plant transformation. The pBECKS2000 series of binary vectors exploits the Cre/ loxP site-specific recombinase system to facilitate the construction of complex T-DNA vectors. The new plasmids enable the rapid generation of T-DNA vectors in which multiple genes are linked, without relying on the availability of purpose-built cassette systems or demanding complex, and therefore inefficient, ligation reactions. The vectors incorporate facilities for the removal of transformation markers from transgenic plants, while still permitting simple in vitro manipulations of the T-DNA vectors. A 'shuttle' or intermediate plasmid approach has been employed. This permits independent ligation strategies to be used for two gene sets. The intermediate plasmid sequence is incorporated into the binary vector through a plasmid co-integration reaction which is mediated by the Cre/loxP site-specific recombinase system. This reaction is carried out within Agrobacterium cells. Recombinant clones, carrying the co-integrative binary plasmid form, are selected directly using the antibiotic resistance marker carried on the intermediate plasmid. This strategy facilitates production of co-integrative T-DNA binary vector forms which are appropriate for either (1) transfer to and integration within the plant genome of target and marker genes as a single T-DNA unit; (2) transfer and integration of target and marker genes as a single T-DNA unit but with a Cre/loxP facility for site-specific excision of marker genes from the plant genome; or (3) co-transfer of target and marker genes as two independent T-DNAs within a single-strain Agrobacterium system, providing the potential for segregational loss of marker genes.

L15 ANSWER 2 OF 4 MEDLINE on STN ACCESSION NUMBER: 93323197 MEDLINE PubMed ID: 8392598 DOCUMENT NUMBER:

TITLE:

Efficient generation of infectious recombinant baculoviruses by site-specific transposon-mediated insertion of foreign genes into a baculovirus genome

propagated in Escherichia coli.

AUTHOR: CORPORATE SOURCE: Luckow V A; Lee S C; Barry G F; Olins P O

Cellular and Molecular Biochemistry, Monsanto Corporate

Research, Chesterfield, Missouri 63198.

Journal of virology, (1993 Aug) 67 (8) 4566-79. SOURCE:

Journal code: 0113724. ISSN: 0022-538X.

United States PUB. COUNTRY:

Journal; Article; (JOURNAL ARTICLE) DOCUMENT TYPE:

English LANGUAGE:

FILE SEGMENT: Priority Journals

199308 ENTRY MONTH:

ENTRY DATE: Entered STN: 19930826 Last Updated on STN: 19990129 Entered Medline: 19930816

AΒ The construction and purification of recombinant baculovirus vectors for the expression of foreign genes in insect cells by standard transfection and plaque assay methods can take as long as 4 to 6 weeks. This period can be reduced to several days by using a novel baculovirus shuttle vector (bacmid) that can replicate in Escherichia coli as a plasmid and can infect susceptible lepidopteran insect cells. The bacmid is a recombinant virus that contains a mini-F replicon, a kanamycin resistance marker, and attTn7, the target site for the bacterial transposon Tn7. Expression cassettes comprising a baculovirus promoter driving expression of a foreign gene that is flanked by the left and right ends of Tn7 can transpose to the target bacmid in E. coli when Tn7 transposition functions are provided in trans by a helper plasmid. foreign gene is expressed when the resulting composite bacmid is introduced into insect cells.

L15 ANSWER 3 OF 4 MEDLINE on STN MEDLINE ACCESSION NUMBER: 88210531 DOCUMENT NUMBER: PubMed ID: 3449223

Terminal segment of Kluyveromyces lactis linear DNA TITLE:

plasmid pGKL2 supports autonomous replication of

hybrid plasmids in Saccharomyces cerevisiae.

Fujimura H; Hishinuma F; Gunge N AUTHOR:

CORPORATE SOURCE: Mitsubishi Kasei Institute of Life Sciences, Tokyo, Japan.

Current genetics, (1987) 12 (2) 99-104. SOURCE: Journal code: 8004904. ISSN: 0172-8083.

United States PUB. COUNTRY:

DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)

English LANGUAGE:

Priority Journals FILE SEGMENT: GENBANK-X05601 OTHER SOURCE:

198806 ENTRY MONTH:

Entered STN: 19900308 ENTRY DATE:

> Last Updated on STN: 19990129 Entered Medline: 19880617

By use of linear DNA plasmid pGKL2 from the yeast Kluyveromyces AΒ lactis we have constructed hybrid plasmids carrying a LEU2 gene of Saccharomyces cerevisiae as a selectable marker. The replication properties of hybrid plasmids in yeasts were investigated. We demonstrated that the insertion of a LEU2 gene into pGKL2 resulted in circularization of the hybrid plasmids and pGKL2 segment supported autonomous replication of the plasmids. Moreover, the hybrid plasmids propagated autonomously, independently of the presence of the natural pGKL2 plasmid.

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on STN

ACCESSION NUMBER: 89010830 EMBASE

1989010830 DOCUMENT NUMBER:

A broad-host-range shuttle system for gene insertion into TITLE:

the chromosomes of Gram-negative bacteria.

Barry G.F. AUTHOR:

Plant Microbiology Group, Biological Sciences Department, CORPORATE SOURCE:

Monsanto Company, St. Louis, MO 63198, United States Gene, (1988) Vol. 71, No. 1, pp. 75-84.

SOURCE:

ISSN: 0378-1119 CODEN: GENED6

COUNTRY: Netherlands Journal DOCUMENT TYPE:

022 Human Genetics FILE SEGMENT:

English LANGUAGE: SUMMARY LANGUAGE: English

Entered STN: 911212 ENTRY DATE:

Last Updated on STN: 911212

A deletion derivative of transposon Tn7 containing the Escherichia coli lacZY genes as a selectable marker for insertion of foreign DNA into the chromosomes of soil bacteria was improved to facilitate the cloning of additional genes and their insertion by this element. This report describes a series of plasmid vectors that enable this cloning to be carried out in small, high-copy, narrow host-range plasmids. The final Tn element can then be easily moved (by transposition) without further use of restriction enzymes, to plasmids suitable for delivering it to the bacterial chromosome. The very high specificity for insertion of $\mathbf{Tn7}$ into single locations in bacterial chromosomes has been exploited in the construction of a shuttle system for delivering these $\mathbf{Tn7}$ elements.

	Document ID	Title
1	US 20050142562 A1	High throughput generation and screening of fully human antibody repertoire in yeast
2	US 20050124010 A1	Whole cell engineering by mutagenizing a substantial portion of a starting genome combining mutations and optionally repeating
3	US 20050123996 A1	Assembly and screening of highly complex and fully human antibody repertoire in yeast
4	US 20050112141 A1	Compositions and methods for treatment of neoplastic disease
5	US 20050059109 A1	Methods and compositions for polypeptide engineering
6	US 20040245317 A1	Artificial chromosomes that can shuttle between bacteria yeast and mammalian cells
7	US 20040219516 A1	Viral vectors containing recombination sites
8	US 20040214783 A1	Compositions and methods for treatment of neoplastic disease
9	US 20040214277 A1	Methods and compositions for polypeptide engineering
10	US 20040067532 A1	High throughput generation and affinity maturation of humanized antibody

	Document ID	Title
11	US 20040005591 A1	Cloning system for construction of recombinant expression vectors
12	US 20030219817 A1	Assembly and screening of highly complex and fully human antibody repertoire in yeast
13	US 20030186356 A1	Methods for generating polynucleotides having desired characteristics by iterative selection and recombination
14	US 20030165990 A1	Generation of highly diverse library of expression vectors via homologous recombination in yeast
15	US 20030157113 A1	Compositions and methods for treatment of neoplastic disease
16	US 20030027213 A1	Haploid yeast cells transformed with a library of expression vectors encoding a fully human antibody repertoire
17	US 20030027156 A1	Methods and compositions for polypeptide engineering
18	US 20020177551 A1	Compositions and methods for treatment of neoplastic disease
19	US 20020051976 A1	METHODS AND COMPOSITIONS FOR POLYPEPTIDE ENGINEERING
20	US 6610472 B1	Assembly and screening of highly complex and fully human antibody repertoire in yeast

	Do	ocument :	ID	Title
21	US	6573098	В1	Nucleic acid libraries
22	US	6551828	R1	Compositions and methods for generating expression vectors through site-specific recombination
23			В1	Methods for generating polynucleotides having desired characteristics by iterative selection and recombination
24	US	6506603	В1	Shuffling polynucleotides by incomplete extension
25	US	6506602	B1	Methods for generating polynucleotides having desired characteristics by iterative selection and recombination
26	US	6413774		Methods for generating polynucleotides having desired characteristics by iterative selection and recombination
27	US	6410271		Generation of highly diverse library of expression vectors via homologous recombination in yeast
28	us	6410246	—	Highly diverse library of yeast expression vectors
29	US	6406863		High throughput generation and screening of fully human antibody repertoire in yeast

	Do	cument :	ID	Title
30	US	6395547	В1	Methods for generating polynucleotides having desired characteristics by iterative selection and recombination
			В1	Methods for generating polynucleotides having desired characteristics by iterative selection and recombination
32	US	6344356	В1	Methods for recombining nucleic acids
33	US	6323030	В1	Methods for generating polynucleotides having desired characteristics by iterative selection and recombination
34	US	6291242	В1	Methods for generating polynucleotides having desired characteristics by iterative selection and recombination
35 ·	US	6180406	B1	Methods for generating polynucleotides having desired characteristics by iterative selection and recombination
36	US	6165793	Α	Methods for generating polynucleotides having desired characteristics by iterative selection and recombination
37	US	6117679	Α	Methods for generating polynucleotides having desired characteristics by iterative selection and recombination



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